

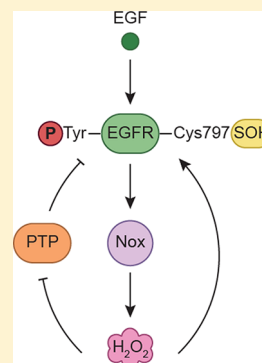
Redox Regulation of Epidermal Growth Factor Receptor Signaling through Cysteine Oxidation

Thu H. Truong[†] and Kate S. Carroll^{*‡}

[†]Department of Chemistry, University of Michigan, Ann Arbor, Michigan 48109, United States

[‡]Department of Chemistry, The Scripps Research Institute, Jupiter, Florida 33456, United States

ABSTRACT: Epidermal growth factor receptor (EGFR) exemplifies the family of receptor tyrosine kinases that mediate numerous cellular processes, including growth, proliferation, and differentiation. Moreover, gene amplification and EGFR mutations have been identified in a number of human malignancies, making this receptor an important target for the development of anticancer drugs. In addition to ligand-dependent activation and concomitant tyrosine phosphorylation, EGFR stimulation results in the localized generation of H₂O₂ by NADPH-dependent oxidases. In turn, H₂O₂ functions as a secondary messenger to regulate intracellular signaling cascades, largely through the modification of specific cysteine residues within redox-sensitive protein targets, including Cys797 in the EGFR active site. In this review, we highlight recent advances in our understanding of the mechanisms that underlie redox regulation of EGFR signaling and how these discoveries may form the basis for the development of new therapeutic strategies for targeting this and other H₂O₂-modulated pathways.



Activation of receptor tyrosine kinases (RTKs) by their respective extracellular ligands (e.g., growth factors) initiates signaling cascades that regulate cellular proliferation, differentiation, migration, and survival.¹ Among the 58 RTKs identified in the human genome, epidermal growth factor (EGF) receptor (EGFR) has served as the quintessential model for understanding RTK biology in physiological signaling and cancer. EGFR, also known as HER1 (or erbB1), is a transmembrane protein grouped into a subfamily that consists of three additional, closely related receptors: HER2 (erbB2), HER3 (erbB3), and HER4 (erbB4). EGFR is comprised of a glycosylated extracellular ligand-binding domain, a transmembrane domain, and an intracellular domain containing its tyrosine kinase core. In response to ligand (e.g., EGF) binding, EGFR forms homo- or heterodimers with other HER family members, followed by autophosphorylation of key tyrosine residues located within the tyrosine kinase domain.² Once activated, EGFR relays the signal through a variety of downstream intracellular signaling cascades, including the Ras/mitogen-activated protein kinase (MAPK) pathway or the phosphatidylinositol 3'-kinase (PI3K)/Akt pathway. Since its discovery,^{3–6} EGFR has been widely studied with regard to its physiologic and pathological settings. In particular, EGFR and related family members have been found to be mutated or amplified in a number of human lung and breast cancers, making them attractive targets for the development of therapeutics.^{7–9}

Beginning in the 1990s, data from several research groups demonstrated that binding of EGF to EGFR triggered the production of endogenous hydrogen peroxide (H₂O₂) in cells (Figure 1).^{10,11} Moreover, it was also established that other growth factors (FGF,¹² PDGF,¹³ VEGF,¹⁴ and insulin¹⁵), cytokines (TNF- α ^{12,16} and interleukin-1¹⁶), and angiotensin

II¹⁷ stimulate H₂O₂ generation. Although chronic exposure to high concentrations of H₂O₂ can lead to a cellular condition known as oxidative stress, cells can also utilize H₂O₂ as a secondary messenger to regulate physiological signal transduction.^{18–21} H₂O₂ modulates signaling pathways largely through the modification of specific cysteine residues located within redox-sensitive protein targets (Figure 2A). The direct product of the reaction between H₂O₂ and a protein thiolate (RS[–]) is sulfenic acid (RSOH). Such protein “sulfenylation” can be reversed by thiol-disulfide oxidoreductases of the thioredoxin (Trx) superfamily throughout the intracellular milieu and, thus, constitutes a facile switch for modulating protein function, akin to phosphorylation. Associated regulatory redox modifications of cysteine include sulfenamides, nitrosothiols, disulfides, sulfinic acids, and sulfonic acids (Figure 2B). Over the past several years, an increasing number of studies have demonstrated important functional roles for protein sulfenic acids in cell signaling.^{22–26} In particular, kinases and phosphatases are both known to undergo cysteine-based redox regulation,^{27,28} and the collective efforts of many researchers have established that these enzymes are regulated by endogenous H₂O₂ produced during EGF-mediated cell signaling.²⁹

In this review, we present a historical overview of EGFR and ligand-mediated production of H₂O₂ as well as the molecular mechanisms involved in redox regulation of this signaling pathway (Figure 1). We begin by highlighting early studies linking EGFR activation to EGF-induced H₂O₂ production. Subsequently, we address the effects of redox modulation on

Received: October 22, 2012

Revised: November 26, 2012

Published: November 27, 2012

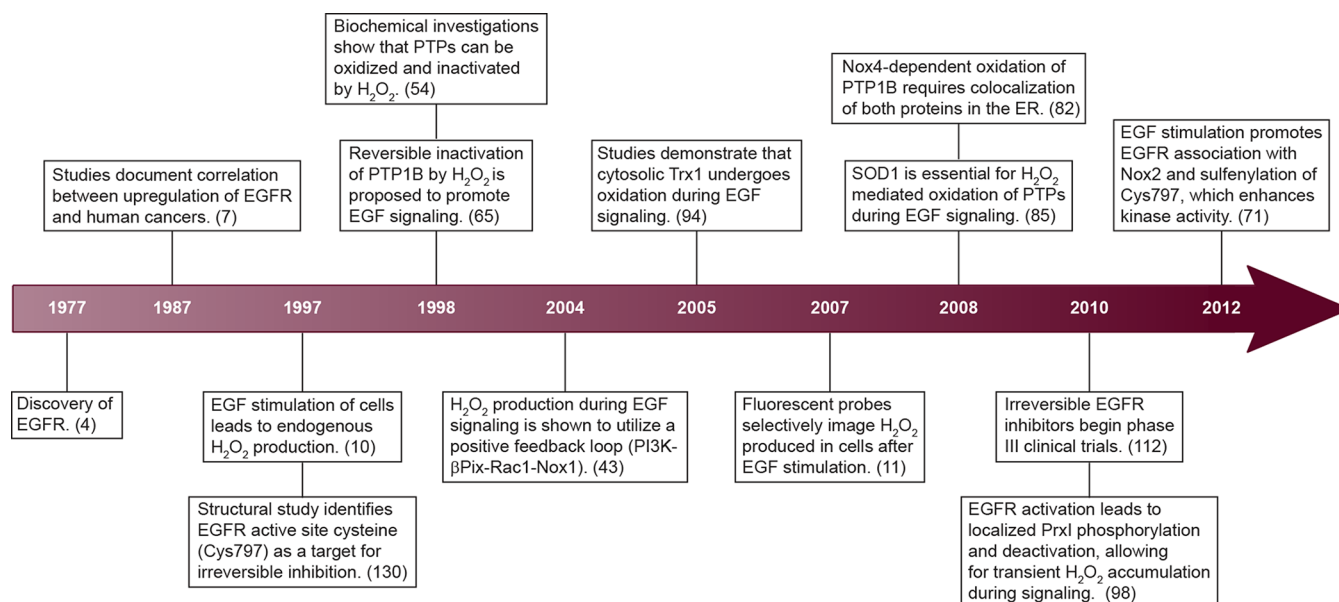


Figure 1. Timeline outlining key events and discoveries relating to redox regulation of EGFR signaling through cysteine oxidation.

two major pathways downstream of EGFR, Ras/MAPK and PI3K/Akt. Additionally, we discuss the interplay between H_2O_2 -mediated inactivation of protein tyrosine phosphatases (PTPs) and kinase activation on net cellular levels of tyrosine (Tyr) phosphorylation. Key enzymes involved in the generation and metabolism of H_2O_2 within EGFR signaling pathways will also be covered. Finally, we focus on recent examples from the literature demonstrating direct oxidation and regulation of EGFR by H_2O_2 and how these discoveries may form the basis for the development of new therapeutic strategies for targeting this and other H_2O_2 -modulated pathways.

■ EARLY EVIDENCE OF EGF-MEDIATED H_2O_2 PRODUCTION AND REDOX REGULATION OF EGFR SIGNALING

In 1995, Gamou and Shimizu published the first report to suggest a connection between H_2O_2 and EGFR. In that study, they examined the effect of exogenously added H_2O_2 on EGFR phosphorylation.³⁰ EGFR-hyperproducing human squamous carcinoma NA cells treated with H_2O_2 (0–1 mM) exhibited an increase in the level of incorporation of [^{32}P]phosphate, albeit at half the signal observed for EGF-stimulated cells. On the basis of data obtained from tryptic phosphopeptide mapping, this discrepancy was attributed to the fact that H_2O_2 might preferentially enhance EGFR Tyr phosphorylation, whereas EGF stimulation would trigger both serine/threonine (Ser/Thr) and Tyr receptor phosphorylation.

During this same time frame (i.e., the mid-1990s), H_2O_2 molecules expanded beyond the traditional view as “toxic byproducts of aerobic metabolism” and began to emerge as secondary messengers in physiological cell signaling. For H_2O_2 to serve as a signaling molecule, its concentration must increase rapidly above the steady-state threshold (i.e., high nanomolar to low millimolar) and remain elevated long enough for it to oxidize protein effectors.³¹ In one of the earliest examples, platelet-derived growth factor (PDGF) was shown to induce endogenous H_2O_2 generation, which is correlated with enhanced Tyr phosphorylation, activation of MAPK pathways,

DNA synthesis, and chemotaxis.¹³ These results suggested that H_2O_2 might act as a signaling molecule generated in response to growth factor stimulation. Soon thereafter, Rhee and colleagues reported that addition of EGF to EGFR-over-expressing human epidermoid carcinoma A431 cells significantly elevated levels of intracellular reactive oxygen species (ROS)¹⁰ as measured by 2',7'-dichlorofluorescein diacetate (DCFH-DA).³² Enzymes such as catalase, peroxiredoxins (Prxs), and glutathione peroxidases (Gpxs) scavenge endogenous H_2O_2 by catalyzing the dismutation (catalase) or reduction (Prxs, Gpxs) of H_2O_2 .^{33,34} Introduction of exogenous catalase in EGF-stimulated A431 cells by electroporation attenuated the intracellular buildup of ROS, suggesting that H_2O_2 was the major ROS involved in EGF-dependent signal transduction.¹⁰ EGF-induced increases in levels of Tyr phosphorylation of PLC- γ 1, a well-characterized physiological substrate of EGFR, were also blunted by catalase incorporation. Although the precise role and/or target(s) of H_2O_2 generated for EGFR signaling was not directly addressed in this study, the authors proposed that inhibition of cysteine-dependent PTPs by H_2O_2 may be required to increase the steady-state level of protein Tyr phosphorylation. This landmark contribution by Rhee and co-workers set the stage for delineating the molecular details underlying redox regulation of EGFR signaling.

In a separate study, Goldkorn et al. reported that exogenous H_2O_2 stimulated EGFR tyrosine kinase activity and increased receptor half-life.³⁵ Experiments performed in A549 human lung adenocarcinoma epithelial cells and isolated membrane fractions showed an increase in EGFR Tyr autophosphorylation levels when treated with H_2O_2 (0–200 μ M) and also markedly enhanced receptor activation in conjunction with the native EGF ligand. Pulse–chase experiments with [^{35}S]methionine revealed that the EGFR half-life was \sim 8 h when treated with EGF, whereas H_2O_2 treatment extended the receptor half-life to 18 h (combined treatment with EGF and H_2O_2 yielded a half-life of 12 h). Additionally, two-dimensional phosphoamino acid analysis corroborated earlier findings³⁰ that the phosphorylation site distribution was shifted predominantly toward Tyr after exposure to H_2O_2 . On the basis of these data, the authors of this study postulated that EGF- and H_2O_2 -induced receptor

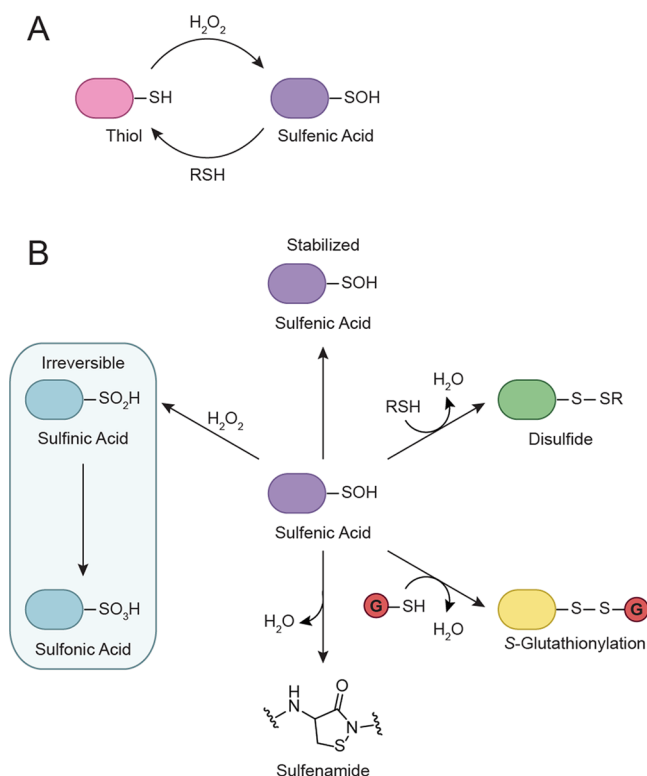


Figure 2. Oxidative modification of cysteine residues by hydrogen peroxide (H₂O₂). (A) The initial reaction product of a thiolate with H₂O₂ yields sulfenic acid (RSOH). This modification, also known as sulfonylation, is reversible and can be directly reduced back to the thiol form or indirectly through disulfide bond formation. (B) Sulfenic acids can be stabilized by the protein microenvironment and/or undergo subsequent modification. For example, they can condense with a second cysteine in the same protein or a different protein to generate a disulfide bond. Alternatively, reaction with the low-molecular weight thiol glutathione (GSH, red circle) affords a mixed disulfide through a process known as S-glutathionylation. In a few proteins, such as PTP1B, nucleophilic attack of a backbone amide on RSOH results in sulfenamide formation. Sulfonyl groups can also oxidize further to the sulfinic (RSO₂H) and/or sulfonic (RSO₃H) acid form under conditions of high oxidative stress.

activation may have separate functions and represent an alternate mechanism by which EGFR signaling can be tuned in parallel to treatment with its native ligand.

■ EFFECT OF H₂O₂ ON SIGNALING NETWORKS DOWNSTREAM OF EGFR

After ligand binding, EGFR transmits activation signals to prominent downstream cascades, including the Ras/MAPK and PI3K/Akt pathways (Figure 3A). In addition to the overall increase in the level of Tyr autophosphorylation of EGFR, the endogenous generation of H₂O₂ also appears to modulate the activity of these two signaling routes. Receptor activation initiates the Ras-Raf-MEK-Erk1/2 signaling module through recruitment of Src homology 2 domain-containing (SHC) adaptor protein, growth factor receptor-bound protein 2 (Grb2), and a guanine nucleotide exchange protein (SOS) to form the SHC-Grb2-SOS complex, a process stimulated by elevated intracellular H₂O₂ levels.³⁶ Once associated with the receptor, SOS facilitates guanine nucleotide exchange to activate Ras, which subsequently activates a kinase cascade that includes Raf (MAP3K), MEK (MAP2K), and Erk1/2

(MAPK). Oxidative stress is known to influence the MAPK signaling pathways, but little is known about the molecular mechanisms responsible for such effects.³⁷ For example, Erk1/2 is activated in response to exogenous H₂O₂, which enhances cell survival after oxidant injury.^{38,39} On the other hand, it is not known whether the activity of Erk1/2 kinase is directly modulated by ROS.

EGFR also transmits signals through the PI3K/Akt pathway.⁴⁰ In response to EGF stimulation, PI3K increases the levels of phosphatidylinositol 3,4,5-trisphosphate (PIP₃), which leads to recruitment and activation of serine/threonine kinase, Akt at the plasma membrane.^{41,42} The activity of the PI3K/Akt pathway is balanced by action of the opposing lipid phosphatase, PTEN, which will be discussed in later sections of this review. In cells overexpressing NADPH oxidase isoform Nox1, an increased level of production of endogenous H₂O₂ elevates intracellular PIP₃ levels and consequently abrogates the ability of PTEN to promote downstream signaling.⁴³ EGFR-dependent activation of Akt has also been shown to enhance cell survival during oxidative stress-induced apoptosis, analogous to Erk1/2.⁴⁴ Interestingly, structural analysis by X-ray crystallography has revealed that Akt2 can form an intramolecular disulfide bond between two cysteines in the activation loop, which inhibits the kinase.⁴⁵ Overexpression of glutaredoxin (Grx) has been shown to protect Akt against H₂O₂-induced oxidation, resulting in sustained phosphorylation of Akt and inhibition of apoptosis.⁴⁶ A more recent study is indicative of isoform-specific regulation of Akt by PDGF-induced H₂O₂.⁴⁷ In particular, the authors demonstrate that Akt2 Cys124 undergoes sulfenic acid modification during growth factor signaling, which inactivates the kinase.

EGFR activation can also occur through a process known as transactivation. In this scenario, ligand-inaccessible RTKs may still initiate downstream signaling in lieu of ligand-receptor-induced activation. Production of H₂O₂ induced by other ligands such as angiotensin II activates c-Src, a redox-regulated kinase.⁴⁸ In turn, EGFR undergoes activation through c-Src-initiated receptor Tyr phosphorylation to propagate downstream signaling.^{48–50} Neighboring EGFR kinases may then undergo activation in a lateral-based mechanism. Transactivation represents another level by which EGFR activity can be modulated by H₂O₂. In addition, other studies have implicated oxidative inactivation of PTPs in promoting EGFR transactivation.⁵¹

■ INTERPLAY OF REVERSIBLE TYROSINE PHOSPHORYLATION AND REDOX-DEPENDENT SIGNALING

Regulation of tyrosine phosphorylation depends on the delicate balance between the action of protein kinases and phosphatase (Figure 3B,C). In EGFR signaling, the coordinated regulation of this equilibrium allows for rapid response to changing growth factor levels, whereas dysregulated kinase and phosphatase activity can have severe pathological consequences such as cancer, diabetes, and inflammation.^{52,53} Because the balance of these two opposing forces is a central event in cell signaling, it is not surprising that phosphatase as well as kinase activities are tightly regulated at several levels, including cysteine-based redox modulation. In large part, because of early studies by Denu and Tanner,⁵⁴ attention rapidly focused on PTPs as the direct targets of H₂O₂ (Figure 3D). The PTP superfamily contains a signature motif, (I/V)HCXXGXXR(S/T), which includes an invariant cysteine residue that functions

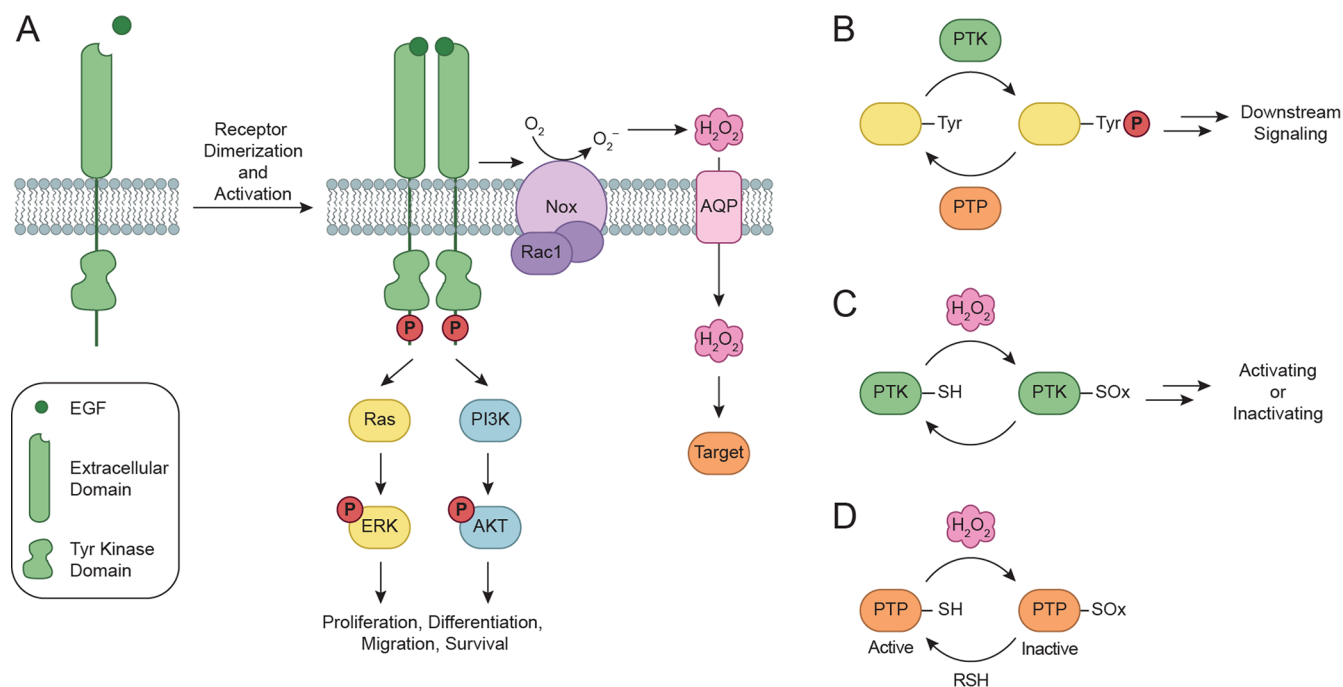


Figure 3. EGFR signaling pathways and general mechanisms for thiol-based redox modulation of signaling proteins. (A) Binding of EGF to EGFR induces receptor dimerization, followed by autophosphorylation of tyrosine (Tyr) residues (red circles) within its cytoplasmic domain. In turn, these phosphorylated Tyr residues serve as docking sites for associating proteins to activate a number of downstream signaling cascades. Two such pathways, Ras/ERK and PI3K/AKT, are shown here for the sake of simplicity. The EGF–EGFR interaction also triggers the assembly and activation of NADPH oxidase (Nox) complexes, followed by subsequent production of H₂O₂ through spontaneous dismutation or action of SOD. Once formed, endogenous H₂O₂ may pass through specific aquaporin (AQP) channels and/or diffuse across the membrane to reach the intracellular cytosol. Transient increases in H₂O₂ levels lead to the oxidation of local redox targets. (B) Model for redox-dependent signal transduction. Protein tyrosine kinases (PTKs) catalyze the transfer of γ -phosphoryl groups from ATP to tyrosine hydroxyls of proteins, whereas protein tyrosine phosphatases (PTPs) remove phosphate groups from phosphorylated tyrosine residues. PTPs function in a coordinated manner with PTKs to control signaling pathways to regulate a diverse array of cellular processes. (C) Regulatory cysteines in protein kinases can undergo oxidation or reduction to modulate their function. Depending on the kinase, redox modifications can stimulate or inhibit function. (D) Oxidation of the conserved active site cysteine residue in PTPs inactivates these enzymes, which can be restored by reducing the oxidized residue to its thiol form. SOx represents oxidized cysteine.

as a nucleophile in catalysis. The catalytic cysteine of PTPs is characterized by a low pK_a value ranging from 4.6 to 5.5 because of the unique electrostatic environment of the active site, which also renders the enzyme susceptible to inactivation by reversible oxidation.^{55,56} Second-order rate constants for oxidation of the PTP cysteine thiolate by H₂O₂ have been measured in vitro and range from 10 to 160 M⁻¹ s⁻¹.^{54,57,58}

Protein tyrosine phosphatase 1B (PTP1B) functions as a negative regulator of the EGFR signaling pathway by directly targeting phosphorylated tyrosine residues that control signaling output.^{59–62} Given that PTP1B is localized exclusively to the cytoplasmic face of the endoplasmic reticulum (ER), it has been proposed to dephosphorylate activated EGFR at sites of contact between the ER and plasma membranes or upon trafficking of internalized EGFR in the proximity of the ER.^{63,64} Studies reported in 1998 provided the first indication that EGF-mediated H₂O₂ production is correlated with oxidation and inactivation of PTP1B.⁶⁵ The level of incorporation of radiolabeled iodoacetic acid (IAA), a sulfhydryl-modifying reagent that reacts with active site Cys215 of PTP1B,⁵⁴ was decreased in PTP1B after EGF stimulation of A431 cells, consistent with oxidation of this essential residue. Conversely, treatment with dithiothreitol (DTT), thioredoxin (Trx), or glutaredoxin (Grx) as a reductant readily reversed PTP1B inhibition. Interestingly, assays with recombinant protein indicated that the Trx system functioned more efficiently as

an electron donor for PTP1B reactivation, as compared to Grx or glutathione (GSH), suggesting that Trx may function as the physiological reductant.⁶⁵

The phosphatase and tensin homologue (PTEN) is another PTP known to maintain a closely intertwined relationship with EGFR. PTEN exhibits dual protein and lipid phosphatase activity and functions as a negative regulator of the PI3K/Akt signaling pathway,⁶⁶ one of the two major signaling routes downstream of EGFR. PTEN contains five cysteine residues in its catalytic domain and undergoes reversible inactivation by H₂O₂.⁶⁷ Site-directed mutagenesis and mass spectrometry indicated that Cys124 was the primary target of H₂O₂, yielding a sulfenic acid intermediate that condenses with Cys71 to form an intramolecular disulfide bond.⁶⁸ Cellular studies also point to a connection between growth factor-induced generation of H₂O₂ and reversible inactivation of PTEN. For example, EGF stimulation of cells results in elevated levels of oxidized PTEN in lysates, as indicated by an electrophoretic mobility shift assay that reports on disulfide formation.⁶⁹ In an alternative approach, protein thiols are alkylated by *N*-ethylmaleimide (NEM) in lysates generated from growth factor-stimulated cells. Reversibly oxidized protein thiols are then reduced with DTT and alkylated with biotin-conjugated maleimide. This method was used to examine reversible oxidation of PTEN in response to EGFR activation.⁶⁹ Together, these studies indicate that EGF-induced activation of PI3K correlates with

inactivation of the opposing phosphatase, PTEN. In this way, PTEN inactivation by endogenous H_2O_2 serves as a positive feedback loop to enhance PIP_3 accumulation and/or Akt activation during EGFR signaling.

SHP2 and DEP-1 represent two additional phosphatases that have been shown to interact with EGFR. SHP2 directly associates with EGFR through its SH2 domains and regulates interactions of the receptor with downstream signaling components such as Ras.⁷⁰ Colocalization of SHP2 with EGFR occurs at low nanomolar concentrations (4 ng/mL) of EGF, and SHP2 was identified as the most sensitive PTP in response to signal-derived H_2O_2 .⁷¹ It is unknown if DEP-1 colocalizes with EGFR intracellularly; however, evidence implicates in vitro oxidation of DEP-1.⁷²

■ GENERATION AND METABOLISM OF H_2O_2 DURING EGFR SIGNALING

The family of NADPH oxidase (Nox) enzymes and their dual oxidase counterparts (Duox) generate superoxide by transferring electrons from cytosolic nicotinamide adenine dinucleotide phosphate (NADPH) to molecular oxygen.⁷³ Once it is generated, superoxide is dismutated spontaneously ($\sim 10^5 \text{ M}^{-1} \text{ s}^{-1}$ at pH 7) or enzymatically by superoxide dismutase (SOD; $\sim 10^9 \text{ M}^{-1} \text{ s}^{-1}$) to H_2O_2 and molecular O_2 .^{74,75} The prototypical Nox isoform, Nox2 (also known as gp91^{phox}), was originally identified and characterized in macrophages and neutrophils, where it functions as an integral part of the innate immune system.⁷⁶ The active form of Nox2 exists as a multisubunit complex, consisting of the membrane-bound cytochrome b_{558} (gp91^{phox} and p22^{phox}), several cytosolic proteins (p47^{phox}, p40^{phox}, and p67^{phox}), and the small GTPase Rac1. Since the initial discovery of Nox2, other Nox homologues (Nox1–5 and Duox1 and -2) have been identified in almost every cell type, localized both to the plasma membrane (where they produce superoxide extracellularly) and to intracellular organelles, and serve as major sources of H_2O_2 for signaling.^{77–79}

Several Nox isoforms play a critical role in EGFR-mediated signaling cascades. For example, Park et al. demonstrated that the PI3K pathway constitutes a positive feedback loop for Nox1 activation in growth factor-stimulated cells.⁴³ This study demonstrated that the Rac-guanine nucleotide exchange factor (GEF) β Pix is required for and also augments EGF-induced generation of H_2O_2 . In this loop, β -Pix and activated Rac1 bind to the C-terminal region of Nox1, relieving autoinhibitory constraints. Independent studies have also shown that phosphorylation of Nox activator 1 (NOXA1) on Ser282 by Erk1/2 kinases and on Ser172 by protein kinases C and A weakens the binding of Rac1 to downregulate Nox1.^{80,81}

Nox4 is an isoform that regulates the activity of internalized EGFR. Keaney and colleagues report that Nox4 is localized to the ER in vascular endothelial cells where it appears to regulate the activity of PTP1B in a spatially dependent manner.⁸² Nox4-dependent oxidation of PTP1B required colocalization of both proteins in the ER, as shown by targeting PTP1B to the cytoplasm. Importantly, the study also demonstrated that Nox4-dependent oxidation and inactivation of PTP1B are correlated with a reduced level of phosphorylation of EGFR in the proximity of the ER. The significance of colocalization was also verified by ER targeting of the antioxidant enzyme, catalase. Lastly, EGF stimulation of A431 cells leads to the formation of a specific complex between Nox2 and EGFR, as demonstrated by co-immunoprecipitation experiments.⁷¹

Superoxide dismutase 1 (SOD1) is another enzyme involved in redox mediation of growth factor signaling. SOD1 is an abundant copper/zinc enzyme located in the cytoplasm and belongs to the SOD family of enzymes. Other members of this family include mitochondrial SOD2 (manganese) and extracellular SOD3 (copper/zinc). Although superoxide can spontaneously dismutate to form H_2O_2 , the second-order rate constant of the reaction is enhanced >10000-fold by SOD.^{74,75} Indeed, SODs catalyze the conversion of superoxide into H_2O_2 and molecular O_2 to maintain superoxide at a low steady-state concentration ($\sim 10^{-10} \text{ M}$).^{83,84} Inhibition of SOD1 by the tetrathiomolybdate inhibitor, ATN-224, increases the intracellular concentration of superoxide at the expense of H_2O_2 production, thereby attenuating EGFR and Erk1/2 phosphorylation.⁸⁵

Membrane transport represents another important mechanism for modulating endogenous H_2O_2 produced during EGFR signaling. Early studies demonstrated that EGF stimulation of cells leads to a rapid increase in the concentration of extracellular H_2O_2 and that the addition of catalase to culture medium was sufficient to inhibit EGFR autophosphorylation.^{86,87} These findings raise the question of how extracellularly generated H_2O_2 could mediate intracellular signaling pathways. Although it had been largely assumed that H_2O_2 could diffuse across cellular membranes, more recent evidence indicates that H_2O_2 may preferentially enter the cell through specific plasma membrane aquaporin channels.^{88–90} Collectively, these studies suggest that the number or type of aquaporins expressed on the cell surface might modulate the level of intracellular H_2O_2 available for signaling.

Efforts have also focused on delineating mechanisms for regulating intracellular H_2O_2 levels during EGF-mediated signaling. In this regard, superoxide dismutases, catalase, and other peroxidases all function to protect cells against undue oxidative stress. Prxs are a family of thiol-based peroxidases that catalyze the dismutation of H_2O_2 into water and molecular oxygen.⁹¹ Prx possesses two cysteines that transiently oxidize to form a disulfide as it metabolizes H_2O_2 . The disulfide in Prx is subsequently reduced by the protein disulfide reductase, Trx, to complete the catalytic cycle. Overexpression of the PrxII isoform reduces EGF-induced intracellular H_2O_2 levels.⁹² Another study demonstrated that PrxII overexpression is associated with a decrease in the level of EGF-induced cellular PIP_3 , whereas a dominant negative (DN) form of PrxII increased PIP_3 levels, presumably by H_2O_2 modulation of the PTEN redox state.⁶⁹ Similarly, overexpression of another antioxidant enzyme, glutathione peroxidase-1 (Gpx1), decreased the level of Tyr phosphorylation of EGFR, activation of Akt, and cellular proliferation.⁹³ In addition to SODs and peroxidases, cells rely on the glutaredoxin/glutathione (Grx/GSH), Trx/thioredoxin reductase (Trx/TrxR), and glutathione/glutathione reductase (GSH/GSR) buffering systems. For example, EGFR signaling is associated with subcellular compartmental oxidation of Trx1. Jones and colleagues have measured the redox states of cytosolic and nuclear Trx1 and mitochondrial Trx2 using redox Western blot methods during endogenous H_2O_2 production induced by EGF signaling.⁹⁴ Interestingly, results from this study showed that only the cytoplasmic Trx1 pool undergoes significant oxidation in response to growth factor treatment. Furthermore, the GSH/GSSG redox couple, which was also examined in this study, did not undergo oxidation. This work suggests that physiological H_2O_2 generation in response to EGF signaling is specifically

associated with oxidation of the Trx1 system and not the GSH system. However, whether Trx1 oxidation is part of the signaling mechanism itself or simply results from peroxidase-dependent termination of the redox signal remains an active area of investigation.

The estimated intracellular steady-state concentration of H_2O_2 hovers in the low nanomolar to low micromolar range.³¹ Then again, these estimates assume that H_2O_2 is uniformly distributed throughout the cell. Given that the source of H_2O_2 produced for EGF signaling (e.g., Nox enzymes) is localized to specific regions of the cell, it stands to reason that signal-mediated changes in H_2O_2 concentration may not be homogeneous throughout the cell. Rather, the oxidant concentration near a source of generation must achieve high local concentrations to function effectively as a second messenger. A growing body of research has focused on delineating the mechanisms that facilitate localized increases in intracellular H_2O_2 levels during growth factor signaling. Although the cell contains millimolar concentrations of GSH, it reacts too slowly with H_2O_2 to provide much buffering capacity.⁹⁵ By contrast, Prxs are extremely efficient at H_2O_2 elimination, reducing H_2O_2 with second-order rate constants of 10^5 – $10^8 \text{ M}^{-1} \text{ s}^{-1}$.^{96,97} Recent work reported by the Rhee laboratory has shown that membrane-localized PrxI can be deactivated by phosphorylation in EGF-stimulated cells and in mice during wound healing.⁹⁸ Knockdown experiments suggested that c-Src kinase is at least partially responsible for PrxI phosphorylation. RTK activation (e.g., PDGFR) can also lead to overoxidation of the PrxII isoform catalytic cysteine to sulfinic acid, resulting in a transiently inactivated protein.⁹⁸ Collectively, these studies demonstrate that selective inactivation of PrxI and PrxII allows for transient H_2O_2 accumulation around plasma membranes where signaling components are concentrated, while simultaneously preventing toxic accumulation of ROS elsewhere in the cell during EGF signaling (Figure 4). Other mechanisms for modulating localized redox buffering capacity surely await discovery.

■ REGULATION OF INTRINSIC EGFR TYROSINE KINASE ACTIVITY THROUGH CYSTEINE OXIDATION

As outlined above, a growing body of evidence demonstrates that EGFR activity and downstream signaling pathways are regulated by redox-based mechanisms. Up until this point, we have only considered downstream events that enhance the overall extent of EGFR activation, such as PTP inactivation. We have also highlighted key regulatory themes in redox signaling, including colocalization of sources and/or targets of H_2O_2 and modulation of local redox buffering capacity. Until recently, there was scant evidence to indicate that the intrinsic tyrosine kinase activity of EGFR itself might be regulated by endogenous H_2O_2 produced during cell signaling.^{99–101} In the section below, we recount recent work from our own laboratory, which has demonstrated that EGFR is directly modulated by endogenous H_2O_2 , as well as studies from other groups that hint at the possibility of modification by reactive nitrogen species (RNS).

Previous data from our group demonstrated that breast cancer cells associated with an increased level of expression of EGFR are correlated with a substantial increase in the level of protein sulenylation.¹⁰² Prompted by this observation, we conducted a more detailed investigation of EGF-induced protein sulenylation in A431 cells.⁷¹ Facilitating these studies,

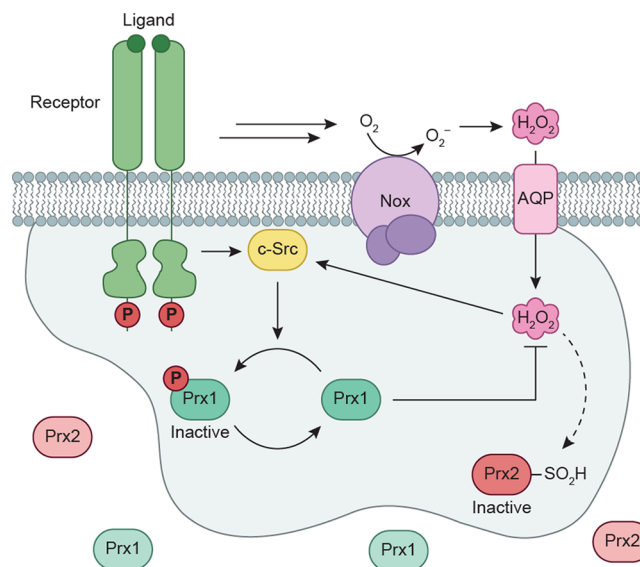


Figure 4. Redox regulation of peroxiredoxins (Prxs) during EGF signaling. Receptor activation results in localized phosphorylation and inactivation of peroxiredoxin I (PrxI) by PTKs, such as the redox-regulated cytoplasmic Src (c-Src). Deactivation of PrxI diminishes the redox buffering capacity adjacent to the cell membrane, allowing for a transient and localized increase in H_2O_2 levels for signal transduction. Additionally, elevated H_2O_2 concentrations can inactivate Prx2 by oxidation of its catalytic cysteine to sulfinic acid.

we have recently described a new technology that allows for the detection and visualization of sulenylation proteins within intact cells,^{103–106} thereby circumventing concerns associated with the analysis of lysates and/or homogenates, including limited spatiotemporal resolution and oxidation artifacts inherent to the lysis procedure.¹⁰⁷ Inspired by earlier work demonstrating the selective reaction of 5,5-dimethyl-1,3-cyclohexanedione (commercially known as dimedone) with protein sulenic acids¹⁰⁸ (Figure 5A), we developed a suite of bifunctional probes that contain a membrane-permeable analogue of dimedone coupled to an azide or alkyne chemical handle (Figure 5B). An orthogonally functionalized biotin or fluorescent tag can be appended post-cell labeling for detection via the Staudinger ligation¹⁰⁹ or Huisgen [3+2] cycloaddition also known as click chemistry.¹¹⁰ Overall, this general approach provides a facile method for monitoring protein sulenylation directly in living cells (Figure 5C).

Employing this method, we demonstrated that EGFR undergoes sulenylation in response to addition of EGF, even at low nanomolar concentrations of growth factor.⁷¹ (Figure 6A). Reciprocal immunoprecipitation analysis also showed that EGFR and Nox2 became associated in an EGF-dependent fashion.⁷¹ The intracellular kinase domain of EGFR contains six cysteine residues, one of which (Cys797) is located in the ATP-binding pocket (Figure 6B) and is conserved among nine additional receptor and nonreceptor tyrosine kinases (Figure 6C).¹¹¹ Given its active site location and conservation, we hypothesized that Cys797 might be preferentially targeted by endogenous H_2O_2 . Of particular relevance, this residue is selectively targeted by irreversible EGFR inhibitors, such as afatinib, extensively used in basic research and clinical trials for breast and non-small-cell lung cancers.^{112,113} Consistent with this proposal, pretreatment of cells with inhibitors that irreversibly modify Cys797 prevented sulenylation of EGFR. Mass spectrometry was subsequently used to verify Cys797 as

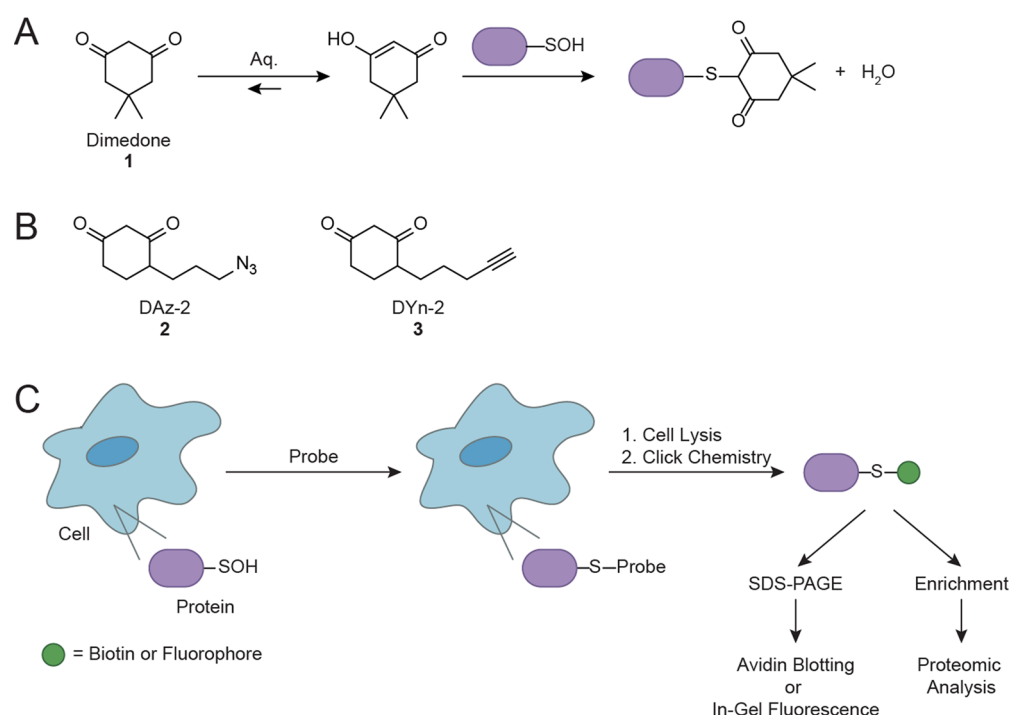


Figure 5. General strategy for detecting protein cysteine sulfenylation (RSOH) in cells. (A) Chemoselective reaction between sulfenic acid and 5,5-dimethyl-1,3-cyclohexanedione (dimedone, 1). (B) Azide- and alkyne-functionalized small-molecule probes for trapping and tagging protein sulfenic acids include DAz-2 (2) and DYn-2 (3). (C) Detection of protein sulfenic acids in living cells. Target cells are incubated with cell-permeable probes to trap and tag protein sulfenic acids in situ. In subsequent steps, lysates are prepared and tagged proteins are further elaborated by attachment of biotin or fluorescence labels via click chemistry that permits detection by Western blot or in-gel fluorescence. Alternatively, biotinylated proteins may be enriched for proteomic analysis.

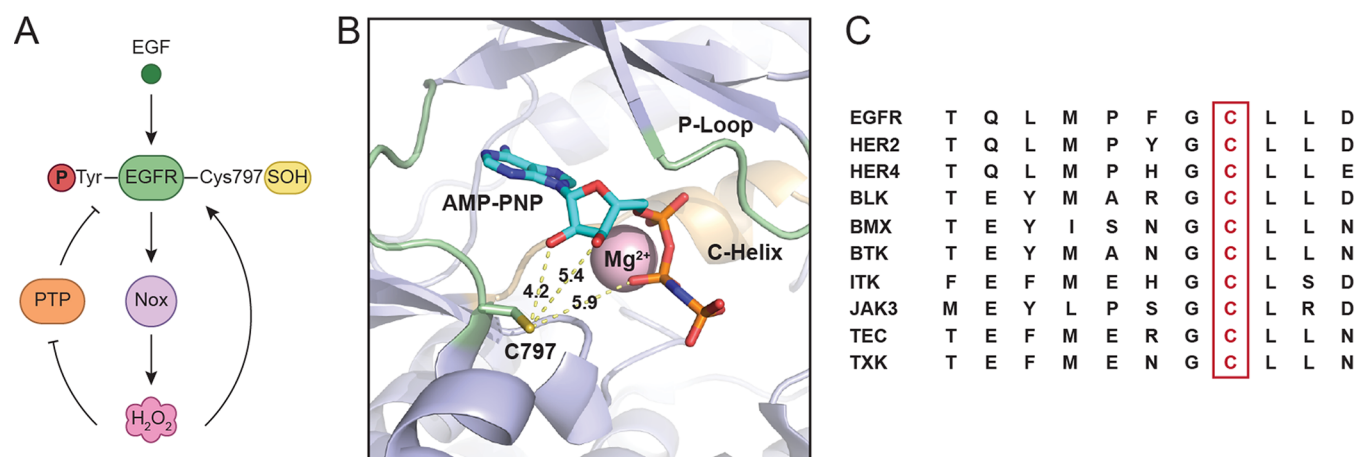


Figure 6. Model for H₂O₂-dependent regulation of EGFR tyrosine kinase activity. (A) Binding of EGF induces production of H₂O₂ through Nox. Nox-derived H₂O₂ directly modifies EGFR cysteine (Cys797) to sulfenic acid in the active site, which enhances its tyrosine kinase activity. Endogenous H₂O₂ can also oxidize and deactivate localized PTPs, leading to a net increase in the level of EGFR phosphorylation. (B) Crystal structure of the EGFR kinase domain (Protein Data Bank entry 3GT8) bound to AMP-PNP, a hydrolysis resistant ATP analogue, and Mg²⁺. Dashed yellow lines and accompanying numbers indicate the distance (angstroms) between the γ -sulfur atom of Cys797 and key substrate functional groups. Note also that Cys797 can adopt different rotamers and sulfenylation of this residue may enhance its ability to participate in electrostatic and hydrogen bonding interactions with its substrate. (C) Abbreviated sequence alignment of EGFR and nine other kinases that harbor a cysteine at the structural position that corresponds to Cys797 (adapted from ref 112).

the specific site of oxidation. Finally, oxidation of Cys797 increased EGFR kinase activity by approximately 5-fold. To put these findings into context, a comparable degree of stimulation is observed for L858R and T790M oncogenic EGFR mutations.^{114–116} Beyond the A431 cell line model, recent studies by our group also indicate that wild-type and several activated mutant EGFR kinases undergo sulfenylation in both

lung and breast tumors (T. H. Truong and K. S. Carroll, unpublished data). Hence, it appears that oxidation of specific residues in PTPs (catalytic Cys) and EGFR (Cys797) both contribute to an increase in the level of downstream signaling (Figure 6A). Current work is directed toward understanding the molecular mechanism by which sulfenylation of EGFR Cys797 enhances kinase activity. The proximity of Cys797 to

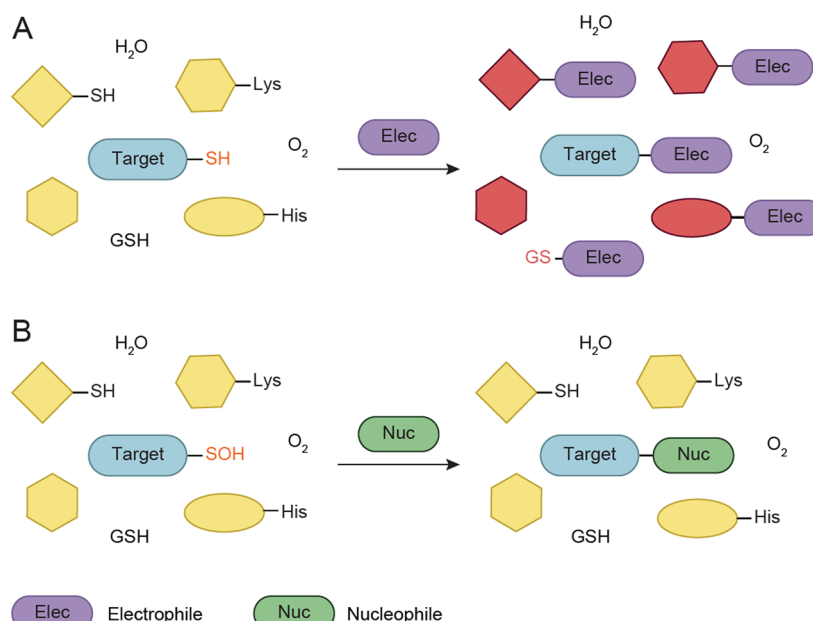


Figure 7. Covalent cysteine-based protein targeting strategies. (A) Conventional covalent inhibitors of kinases inactivate their target through covalent attachment to the cysteine thiol functional group. However, the electrophilic center (e.g., acrylamide, haloacetamide, and vinyl sulfonamide) that reacts with the cysteine can exhibit nonspecific reactivity toward other cellular thiols, including glutathione present at millimolar concentrations inside mammalian cells. The electrophile may also react with other nucleophilic functionalities present in biological systems (amino and imidazole groups of amino acids, various reactive sites in nucleic acid bases, and water). (B) Orthogonal strategy as one potential mechanism for addressing issues associated with employing an electrophilic functional group to target one cysteine among a sea of biological nucleophiles. According to this approach, active site-directed small-molecule inhibitors containing a reactive nucleophilic substituent form a covalent bond with a sulfenic acid-modified cysteine side chain. Such modifications form transiently in specific proteins during H_2O_2 -mediated signal transduction in normal cells but form constitutively in diseases associated with chronically elevated levels of H_2O_2 , including cancer. In the sulfenic acid oxidation state, the electron-deficient sulfur exhibits enhanced electrophilic character that can be selectively targeted by certain nucleophilic compounds. Because sulfenic acid is a unique chemical moiety in biochemistry, this strategy might decrease the potential for off-target activity while retaining the advantages gained by covalent targeting.

the ATP ligand as well as the C-helix and activation segment raises the possibility of transition-state stabilization and/or destabilization of its autoinhibited conformation.¹¹⁶

We have also applied our chemical biology approach to monitor global changes in EGF-dependent protein sulfenylation.⁷¹ Interestingly, addition of growth factor led to widespread changes in protein sulfenylation within cells. EGF induced cysteine oxidation in a dose- and time-dependent manner, which was accompanied by concomitant fluxes in intracellular H_2O_2 levels. Additional experiments showed that treatment with cell-permeable PEGylated catalase attenuated EGF-associated changes in protein sulfenylation, underscoring the importance of endogenous H_2O_2 . Although sulfenylation of EGFR (and several PTPs) was the focus our recent study,⁷¹ many protein targets of EGF-induced endogenous H_2O_2 remain to be identified and will necessitate large-scale proteomic analysis.

Remarkably, a number of studies indicate that EGFR may also undergo modulation by RNS at cysteine residues distinct from Cys797. Reaction of cysteine thiols with RNS, such as nitric oxide (NO), generates nitrosothiols (S-NO). This process, known as S-nitrosylation, is a well-established reversible post-translational cysteine modification and has been implicated in proliferative and antiproliferative cellular effects.¹¹⁷ The approach most often used to identify protein nitrosothiols is known as the biotin switch technique (BST).¹¹⁸ The BST is an indirect method whose success relies heavily on the alkylation of free thiols and the selectivity and/or efficiency of the ascorbate reducing agent toward nitrosothiols.^{119,120}

Nonetheless, this method was utilized in two separate studies reporting S-nitrosylation of EGFR. Treatment of several cell types with an exogenous nitric oxide (NO) donor 1,1-diethyl-2-hydroxy-2-nitrosohydrazine (DEA-NO) at a concentration of 1 mM inhibited EGFR autophosphorylation.^{121,122} Mutation of Cys166 of EGFR, located in the extracellular domain, to serine rendered the receptor resistant to NO-induced inhibition. Alternatively, another cysteine residue located at the extracellular EGFR ligand-binding interface undergoes nitrosylation after exposure to 1 mM exogenous NO donor S-nitroso-L-cysteine (Cys-NO).¹²³ S-Nitrosylation of these residues may inhibit EGFR-mediated signaling by interfering with the ligand interaction site, although this proposal awaits evaluation. By contrast, a more recent study demonstrates that S-nitrosylation induced by the chemical NO donor (Z)-1-[N-(2-aminoethyl)-N-(2-ammonioethyl)amino]diazen-1-ium 1,2-diolate known as DETA-NO can upregulate EGFR signaling and correlates with a transformed breast cancer phenotype.^{124,125} An important goal for future research is to determine whether EGFR undergoes direct S-nitrosylation in response to physiological stimuli.

FUTURE PERSPECTIVES

Of the ~95 receptor and nonreceptor PTKs in the human genome, nine additional members harbor a cysteine residue that is structurally homologous to EGFR Cys797 (Figure 6C), including two EGFR subfamily members, HER2 and HER4. Although this conclusion is speculative at this time, it is possible that this group of kinases is regulated by oxidation of this

residue. EGFR Cys797 and its structural analogues are located at the N-terminal end of an α helix, also known as the N_{cap} position. Interestingly, cysteine is the most sparsely occurring N_{cap} residue in natural proteins, comprising <1% of all these positions.¹²⁶ Interaction of a cysteine located at the N_{cap} position with the helix dipole can drastically lower the thiol pK_a and increases its reactivity.^{127,128} Of note, the N_{cap} effect has also been attributed to the reactivity of the human PrxI catalytic cysteine.⁹⁸ Another subfamily of PTKs, which includes cytoplasmic Src and FGFR1, contains a cysteine located within a glycine rich loop that interacts with the γ -phosphate of ATP.¹²⁹ Oxidation of this residue inhibits the kinase activity of c-Src and FGFR1 in vitro; however, neither kinase has been confirmed as a direct target of endogenous H₂O₂ in cells. More than 150 kinases have a cysteine in or around the nucleotide-binding site, some of which may play similar regulatory roles. However, much more work will be required to define the scope and molecular details underlying the redox-regulated kinome.

EGFR is mutated or amplified in a number of human carcinomas, including breast and lung cancers, which has motivated the development of selective kinase inhibitors, including analogues that covalently modify Cys797 and are currently in phase II and III clinical trials.¹³⁰ The recent findings that EGFR Cys797 undergoes sulfenic acid modification⁷¹ and that elevated EGFR and HER2 levels in cancer cells correlate with a substantial increase in the extent of global protein sulfenylation¹⁰² raise several fundamental questions vis-à-vis cysteine oxidation and thiol-targeted irreversible inhibitors. For example, the acrylamide moiety of irreversible EGFR inhibitors undergoes Michael addition with Cys797 in its thiol form, but these inhibitors would not react with the sulfenic acid or other cysteine chemotypes (Figure 7A). Thus, oxidation of Cys797 could affect the potency of these inhibitors, particularly under conditions of oxidative stress often associated with cancer. On the other hand, the sulfenic acid moiety represents an entirely new opportunity in covalent inhibitor design whereby the electrophilic S atom is targeted using a nucleophilic warhead (Figure 7B). In this approach, the propensity for specific cysteine residues in kinases, and other therapeutically important proteins, to undergo sulfenylation could be exploited for the development of inhibitors that target this unique modification, similar to the proof-of-concept compounds we have recently reported to target oxidized PTPs.¹³¹

CONCLUSIONS

The perspectives presented here highlight the emerging and rapidly expanding role of redox regulation during EGFR signaling. These studies point to the unique chemistry of reactive cysteine residues within specific target proteins, including EGFR. These redox reactions allow covalent regulation of protein function, much like phosphorylation. The expanding array of modifications that target cysteine suggests that we are just beginning to understand the molecular basis for the specificity of redox signaling. One theme that has consistently emerged in numerous studies is the colocalization of the oxidant sources with the redox-regulated target protein. As a case in point, ligand activation triggers the association of EGFR and the NADPH oxidase, Nox2. We have also tried to emphasize the value of selective and cell-permeable chemical approaches to elucidating regulatory mechanisms that govern H₂O₂-mediated sulfenylation of proteins. In addition, we are just beginning to appreciate that different biological oxidants

may target distinct cysteine residues in the same protein and thus lead to unique regulatory outcomes. Given that aberrant sulfenylation of proteins is linked to aggressive cancer phenotypes and that genetic lesions in H₂O₂-metabolizing enzymes can contribute to tumorigenesis, defining mechanisms that control reversible protein sulfenylation will be vital to understanding both human physiology and disease. Finally, it is hoped that the discovery of EGFR as a direct target of H₂O₂ will lead to a broader examination of the redox-regulated kinome and the development of an orthogonal nucleophilic strategy for the covalent inhibition of therapeutically important proteins.

AUTHOR INFORMATION

Corresponding Author

*The Scripps Research Institute, Scripps Florida, 130 Scripps Way, Jupiter, FL 33458. E-mail: kcarroll@scripps.edu. Phone: (561) 228-2460. Fax: (561) 228-2919.

Funding

This work was supported by National Institutes of Health Grant GM102187 (K.S.C.) and funding from the Camille Henry Dreyfus Teacher Scholar Award (K.S.C.).

Notes

The authors declare no competing financial interest.

ABBREVIATIONS

RTK, receptor tyrosine kinase; PTK, protein tyrosine kinase; PTP, protein tyrosine phosphatase; EGF, epidermal growth factor; EGFR, EGF receptor; MAPK, mitogen-activated protein kinase; PI3K, phosphatidylinositol 3'-kinase; PTEN, phosphatase and tensin homologue; PIP₃, phosphatidylinositol 3,4,5-triphosphate; H₂O₂, hydrogen peroxide; ROS, reactive oxygen species; Nox, NADPH oxidase; NADPH, nicotinamide adenine dinucleotide phosphate; SOD, superoxide dismutase; Prx, peroxiredoxin; Gpx, glutathione peroxidase; Grx, glutaredoxin; GSH, glutathione; GSR, glutathione reductase; Trx, thioredoxin; TrxR, thioredoxin reductase; ER, endoplasmic reticulum; DTT, dithiothreitol; IAA, iodoacetic acid; IAM, iodoacetamide; NEM, N-ethylmaleimide; DCFH-DA, 2',7'-dichlorofluorescein diacetate; DN, dominant-negative; RNS, reactive nitrogen species; NO, nitric oxide.

REFERENCES

- (1) Schlessinger, J. (2000) Cell signaling by receptor tyrosine kinases. *Cell* 103, 211–225.
- (2) Schlessinger, J. (2002) Ligand-induced, receptor-mediated dimerization and activation of EGF receptor. *Cell* 110, 669–672.
- (3) Cohen, S. (1962) Isolation of a mouse submaxillary gland protein accelerating incisor eruption and eyelid opening in the new-born animal. *J. Biol. Chem.* 237, 1555–1562.
- (4) Cohen, S., Carpenter, G., and King, L., Jr. (1980) Epidermal growth factor-receptor-protein kinase interactions. Co-purification of receptor and epidermal growth factor-enhanced phosphorylation activity. *J. Biol. Chem.* 255, 4834–4842.
- (5) Carpenter, G., King, L., Jr., and Cohen, S. (1978) Epidermal growth factor stimulates phosphorylation in membrane preparations in vitro. *Nature* 276, 409–410.
- (6) Cohen, S. (2008) Origins of growth factors: NGF and EGF. *J. Biol. Chem.* 283, 33793–33797.
- (7) Macias, A., Azavedo, E., Hagerstrom, T., Klintonberg, C., Perez, R., and Skoog, L. (1987) Prognostic significance of the receptor for epidermal growth factor in human mammary carcinomas. *Anticancer Res.* 7, 459–464.

- (8) Yarden, Y., and Sliwkowski, M. X. (2001) Untangling the ErbB signalling network. *Nat. Rev. Mol. Cell Biol.* 2, 127–137.
- (9) Herbst, R. S. (2004) Review of epidermal growth factor receptor biology. *Int. J. Radiat. Oncol. Biol. Phys.* 59, 21–26.
- (10) Bae, Y. S., Kang, S. W., Seo, M. S., Baines, I. C., Tekle, E., Chock, P. B., and Rhee, S. G. (1997) Epidermal growth factor (EGF)-induced generation of hydrogen peroxide. Role in EGF receptor-mediated tyrosine phosphorylation. *J. Biol. Chem.* 272, 217–221.
- (11) Miller, E. W., Tulyathan, O., Isacoff, E. Y., and Chang, C. J. (2007) Molecular imaging of hydrogen peroxide produced for cell signaling. *Nat. Chem. Biol.* 3, 263–267.
- (12) Lo, Y. Y., and Cruz, T. F. (1995) Involvement of reactive oxygen species in cytokine and growth factor induction of c-fos expression in chondrocytes. *J. Biol. Chem.* 270, 11727–11730.
- (13) Sundaresan, M., Yu, Z. X., Ferrans, V. J., Irani, K., and Finkel, T. (1995) Requirement for generation of H₂O₂ for platelet-derived growth factor signal transduction. *Science* 270, 296–299.
- (14) Colavitti, R., Pani, G., Bedogni, B., Anzevino, R., Borrello, S., Waltenberger, J., and Galeotti, T. (2002) Reactive oxygen species as downstream mediators of angiogenic signaling by vascular endothelial growth factor receptor-2/KDR. *J. Biol. Chem.* 277, 3101–3108.
- (15) May, J. M., and de Haen, C. (1979) Insulin-stimulated intracellular hydrogen peroxide production in rat epididymal fat cells. *J. Biol. Chem.* 254, 2214–2220.
- (16) Meier, B., Radeke, H. H., Selle, S., Younes, M., Sies, H., Resch, K., and Habermehl, G. G. (1989) Human fibroblasts release reactive oxygen species in response to interleukin-1 or tumour necrosis factor- α . *Biochem. J.* 263, 539–545.
- (17) Griendling, K. K., Minieri, C. A., Ollerenshaw, J. D., and Alexander, R. W. (1994) Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. *Circ. Res.* 74, 1141–1148.
- (18) Rhee, S. G. (2006) Cell signaling. H₂O₂, a necessary evil for cell signaling. *Science* 312, 1882–1883.
- (19) D'Autreaux, B., and Toledano, M. B. (2007) ROS as signalling molecules: Mechanisms that generate specificity in ROS homeostasis. *Nat. Rev. Mol. Cell Biol.* 8, 813–824.
- (20) Dickinson, B. C., and Chang, C. J. (2011) Chemistry and biology of reactive oxygen species in signaling or stress responses. *Nat. Chem. Biol.* 7, 504–511.
- (21) Finkel, T. (2011) Signal transduction by reactive oxygen species. *J. Cell Biol.* 194, 7–15.
- (22) Reddie, K. G., and Carroll, K. S. (2008) Expanding the functional diversity of proteins through cysteine oxidation. *Curr. Opin. Chem. Biol.* 12, 746–754.
- (23) Paulsen, C. E., and Carroll, K. S. (2010) Orchestrating redox signaling networks through regulatory cysteine switches. *ACS Chem. Biol.* 5, 47–62.
- (24) Jacob, C., Battaglia, E., Burkholz, T., Peng, D., Bagrel, D., and Montenarh, M. (2012) Control of oxidative posttranslational cysteine modifications: From intricate chemistry to widespread biological and medical applications. *Chem. Res. Toxicol.* 25, 588–604.
- (25) Zheng, M., Aslund, F., and Storz, G. (1998) Activation of the OxyR transcription factor by reversible disulfide bond formation. *Science* 279, 1718–1721.
- (26) Chen, C. Y., Willard, D., and Rudolph, J. (2009) Redox regulation of SH2-domain-containing protein tyrosine phosphatases by two backdoor cysteines. *Biochemistry* 48, 1399–1409.
- (27) Aslan, M., and Ozben, T. (2003) Oxidants in receptor tyrosine kinase signal transduction pathways. *Antioxid. Redox Signaling* 5, 781–788.
- (28) Tonks, N. K. (2005) Redox redux: Revisiting PTPs and the control of cell signaling. *Cell* 121, 667–670.
- (29) Finkel, T. (2012) From sulfenylation to sulfhydration: What a thiolate needs to tolerate. *Sci. Signaling* 5, pe10.
- (30) Gamou, S., and Shimizu, N. (1995) Hydrogen peroxide preferentially enhances the tyrosine phosphorylation of epidermal growth factor receptor. *FEBS Lett.* 357, 161–164.
- (31) Stone, J. R., and Yang, S. (2006) Hydrogen peroxide: A signaling messenger. *Antioxid. Redox Signaling* 8, 243–270.
- (32) Bass, D. A., Parce, J. W., Dechatelet, L. R., Szejda, P., Seeds, M. C., and Thomas, M. (1983) Flow cytometric studies of oxidative product formation by neutrophils: A graded response to membrane stimulation. *J. Immunol.* 130, 1910–1917.
- (33) Giorgio, M., Trinei, M., Migliaccio, E., and Pelicci, P. G. (2007) Hydrogen peroxide: A metabolic by-product or a common mediator of ageing signals? *Nat. Rev. Mol. Cell Biol.* 8, 722–728.
- (34) Halliwell, B., and Gutteridge, J. M. C. (1999) *Free Radicals in Biology and Medicine*, Oxford University Press, Oxford, U.K.
- (35) Goldkorn, T., Balaban, N., Matsukuma, K., Chea, V., Gould, R., Last, J., Chan, C., and Chavez, C. (1998) EGF-Receptor phosphorylation and signaling are targeted by H₂O₂ redox stress. *Am. J. Respir. Cell Mol. Biol.* 19, 786–798.
- (36) Rao, G. N. (1996) Hydrogen peroxide induces complex formation of SHC-Grb2-SOS with receptor tyrosine kinase and activates Ras and extracellular signal-regulated protein kinases group of mitogen-activated protein kinases. *Oncogene* 13, 713–719.
- (37) McCubrey, J. A., Lahair, M. M., and Franklin, R. A. (2006) Reactive oxygen species-induced activation of the MAP kinase signaling pathways. *Antioxid. Redox Signaling* 8, 1775–1789.
- (38) Guyton, K. Z., Liu, Y., Gorospe, M., Xu, Q., and Holbrook, N. J. (1996) Activation of mitogen-activated protein kinase by H₂O₂. Role in cell survival following oxidant injury. *J. Biol. Chem.* 271, 4138–4142.
- (39) Wang, X., Martindale, J. L., Liu, Y., and Holbrook, N. J. (1998) The cellular response to oxidative stress: Influences of mitogen-activated protein kinase signalling pathways on cell survival. *Biochem. J.* 333 (Part 2), 291–300.
- (40) Leslie, N. R. (2006) The redox regulation of PI 3-kinase-dependent signaling. *Antioxid. Redox Signaling* 8, 1765–1774.
- (41) Stokoe, D., Stephens, L. R., Copeland, T., Gaffney, P. R., Reese, C. B., Painter, G. F., Holmes, A. B., McCormick, F., and Hawkins, P. T. (1997) Dual role of phosphatidylinositol-3,4,5-trisphosphate in the activation of protein kinase B. *Science* 277, 567–570.
- (42) Franke, T. F., Kaplan, D. R., Cantley, L. C., and Toker, A. (1997) Direct regulation of the Akt proto-oncogene product by phosphatidylinositol-3,4-bisphosphate. *Science* 275, 665–668.
- (43) Park, H. S., Lee, S. H., Park, D., Lee, J. S., Ryu, S. H., Lee, W. J., Rhee, S. G., and Bae, Y. S. (2004) Sequential activation of phosphatidylinositol 3-kinase, β Pix, Rac1, and Nox1 in growth factor-induced production of H₂O₂. *Mol. Cell. Biol.* 24, 4384–4394.
- (44) Wang, X., McCullough, K. D., Franke, T. F., and Holbrook, N. J. (2000) Epidermal growth factor receptor-dependent Akt activation by oxidative stress enhances cell survival. *J. Biol. Chem.* 275, 14624–14631.
- (45) Huang, X., Begley, M., Morgenstern, K. A., Gu, Y., Rose, P., Zhao, H., and Zhu, X. (2003) Crystal structure of an inactive Akt2 kinase domain. *Structure* 11, 21–30.
- (46) Murata, H., Ihara, Y., Nakamura, H., Yodoi, J., Sumikawa, K., and Kondo, T. (2003) Glutaredoxin exerts an antiapoptotic effect by regulating the redox state of Akt. *J. Biol. Chem.* 278, 50226–50233.
- (47) Wani, R., Qian, J., Yin, L., Bechtold, E., King, S. B., Poole, L. B., Paek, E., Tsang, A. W., and Furdul, C. M. (2011) Isoform-specific regulation of Akt by PDGF-induced reactive oxygen species. *Proc. Natl. Acad. Sci. U.S.A.* 108, 10550–10555.
- (48) Ushio-Fukai, M., Griendling, K. K., Becker, P. L., Hilenski, L., Halleran, S., and Alexander, R. W. (2001) Epidermal growth factor receptor transactivation by angiotensin II requires reactive oxygen species in vascular smooth muscle cells. *Arterioscler., Thromb., Vasc. Biol.* 21, 489–495.
- (49) Chen, K., Vita, J. A., Berk, B. C., and Keaney, J. F., Jr. (2001) c-Jun N-terminal kinase activation by hydrogen peroxide in endothelial cells involves SRC-dependent epidermal growth factor receptor transactivation. *J. Biol. Chem.* 276, 16045–16050.
- (50) Giannoni, E., Buricchi, F., Grimaldi, G., Parri, M., Cialdai, F., Taddei, M. L., Raugei, G., Ramponi, G., and Chiarugi, P. (2008) Redox regulation of anoikis: Reactive oxygen species as essential mediators of cell survival. *Cell Death Differ.* 15, 867–878.

- (51) Reynolds, A. R., Tischer, C., Verveer, P. J., Rocks, O., and Bastiaens, P. I. (2003) EGFR activation coupled to inhibition of tyrosine phosphatases causes lateral signal propagation. *Nat. Cell Biol.* 5, 447–453.
- (52) Tonks, N. K. (2006) Protein tyrosine phosphatases: From genes, to function, to disease. *Nat. Rev. Mol. Cell Biol.* 7, 833–846.
- (53) Blume-Jensen, P., and Hunter, T. (2001) Oncogenic kinase signalling. *Nature* 411, 355–365.
- (54) Denu, J. M., and Tanner, K. G. (1998) Specific and reversible inactivation of protein tyrosine phosphatases by hydrogen peroxide: Evidence for a sulfenic acid intermediate and implications for redox regulation. *Biochemistry* 37, 5633–5642.
- (55) Zhang, Z. Y., and Dixon, J. E. (1993) Active site labeling of the *Yersinia* protein tyrosine phosphatase: The determination of the pKa of the active site cysteine and the function of the conserved histidine 402. *Biochemistry* 32, 9340–9345.
- (56) Lohse, D. L., Denu, J. M., Santoro, N., and Dixon, J. E. (1997) Roles of aspartic acid-181 and serine-222 in intermediate formation and hydrolysis of the mammalian protein-tyrosine-phosphatase PTP1. *Biochemistry* 36, 4568–4575.
- (57) Winterbourn, C. C., and Metodiewa, D. (1999) Reactivity of biologically important thiol compounds with superoxide and hydrogen peroxide. *Free Radical Biol. Med.* 27, 322–328.
- (58) Sohn, J., and Rudolph, J. (2003) Catalytic and chemical competence of regulation of cdc25 phosphatase by oxidation/reduction. *Biochemistry* 42, 10060–10070.
- (59) Flint, A. J., Tiganis, T., Barford, D., and Tonks, N. K. (1997) Development of “substrate-trapping” mutants to identify physiological substrates of protein tyrosine phosphatases. *Proc. Natl. Acad. Sci. U.S.A.* 94, 1680–1685.
- (60) Liu, F., and Chernoff, J. (1997) Protein tyrosine phosphatase 1B interacts with and is tyrosine phosphorylated by the epidermal growth factor receptor. *Biochem. J.* 327 (Part 1), 139–145.
- (61) Wu, Y., Kwon, K. S., and Rhee, S. G. (1998) Probing cellular protein targets of H₂O₂ with fluorescein-conjugated iodoacetamide and antibodies to fluorescein. *FEBS Lett.* 440, 111–115.
- (62) Lou, Y. W., Chen, Y. Y., Hsu, S. F., Chen, R. K., Lee, C. L., Khoo, K. H., Tonks, N. K., and Meng, T. C. (2008) Redox regulation of the protein tyrosine phosphatase PTP1B in cancer cells. *FEBS J.* 275, 69–88.
- (63) Haj, F. G., Markova, B., Klamann, L. D., Bohmer, F. D., and Neel, B. G. (2003) Regulation of receptor tyrosine kinase signaling by protein tyrosine phosphatase-1B. *J. Biol. Chem.* 278, 739–744.
- (64) Haj, F. G., Verveer, P. J., Squire, A., Neel, B. G., and Bastiaens, P. I. (2002) Imaging sites of receptor dephosphorylation by PTP1B on the surface of the endoplasmic reticulum. *Science* 295, 1708–1711.
- (65) Lee, S. R., Kwon, K. S., Kim, S. R., and Rhee, S. G. (1998) Reversible inactivation of protein-tyrosine phosphatase 1B in A431 cells stimulated with epidermal growth factor. *J. Biol. Chem.* 273, 15366–15372.
- (66) Leslie, N. R., Bennett, D., Lindsay, Y. E., Stewart, H., Gray, A., and Downes, C. P. (2003) Redox regulation of PI 3-kinase signalling via inactivation of PTEN. *EMBO J.* 22, 5501–5510.
- (67) Maehama, T., Taylor, G. S., and Dixon, J. E. (2001) PTEN and myotubularin: Novel phosphoinositide phosphatases. *Annu. Rev. Biochem.* 70, 247–279.
- (68) Lee, S. R., Yang, K. S., Kwon, J., Lee, C., Jeong, W., and Rhee, S. G. (2002) Reversible inactivation of the tumor suppressor PTEN by H₂O₂. *J. Biol. Chem.* 277, 20336–20342.
- (69) Kwon, J., Lee, S. R., Yang, K. S., Ahn, Y., Kim, Y. J., Stadtman, E. R., and Rhee, S. G. (2004) Reversible oxidation and inactivation of the tumor suppressor PTEN in cells stimulated with peptide growth factors. *Proc. Natl. Acad. Sci. U.S.A.* 101, 16419–16424.
- (70) Agazie, Y. M., and Hayman, M. J. (2003) Molecular mechanism for a role of SHP2 in epidermal growth factor receptor signaling. *Mol. Cell Biol.* 23, 7875–7886.
- (71) Paulsen, C. E., Truong, T. H., Garcia, F. J., Homann, A., Gupta, V., Leonard, S. E., and Carroll, K. S. (2012) Peroxide-dependent sulfenylation of the EGFR catalytic site enhances kinase activity. *Nat. Chem. Biol.* 8, 57–64.
- (72) Persson, C., Kappert, K., Engstrom, U., Ostman, A., and Sjoblom, T. (2005) An antibody-based method for monitoring in vivo oxidation of protein tyrosine phosphatases. *Methods* 35, 37–43.
- (73) Lambeth, J. D. (2004) NOX enzymes and the biology of reactive oxygen. *Nat. Rev. Immunol.* 4, 181–189.
- (74) McCord, J. M., and Fridovich, I. (1969) Superoxide dismutase. An enzymic function for erythrocuprein (hemocuprein). *J. Biol. Chem.* 244, 6049–6055.
- (75) Hsu, J. L., Hsieh, Y., Tu, C., O'Connor, D., Nick, H. S., and Silverman, D. N. (1996) Catalytic properties of human manganese superoxide dismutase. *J. Biol. Chem.* 271, 17687–17691.
- (76) Babior, B. M., Lambeth, J. D., and Nauseef, W. (2002) The neutrophil NADPH oxidase. *Arch. Biochem. Biophys.* 397, 342–344.
- (77) Suh, Y. A., Arnold, R. S., Lassegue, B., Shi, J., Xu, X., Sorescu, D., Chung, A. B., Griendling, K. K., and Lambeth, J. D. (1999) Cell transformation by the superoxide-generating oxidase Mox1. *Nature* 401, 79–82.
- (78) Cheng, G., Cao, Z., Xu, X., van Meir, E. G., and Lambeth, J. D. (2001) Homologs of gp91phox: Cloning and tissue expression of Nox3, Nox4, and Nox5. *Gene* 269, 131–140.
- (79) De Deken, X., Wang, D., Many, M. C., Costagliola, S., Libert, F., Vassart, G., Dumont, J. E., and Miot, F. (2000) Cloning of two human thyroid cDNAs encoding new members of the NADPH oxidase family. *J. Biol. Chem.* 275, 23227–23233.
- (80) Oh, H., Jung, H. Y., Kim, J., and Bae, Y. S. (2010) Phosphorylation of serine282 in NADPH oxidase activator 1 by Erk desensitizes EGF-induced ROS generation. *Biochem. Biophys. Res. Commun.* 394, 691–696.
- (81) Kroviarski, Y., Debbabi, M., Bachoual, R., Perianin, A., Gougerot-Pocidalo, M. A., El-Benna, J., and Dang, P. M. (2010) Phosphorylation of NADPH oxidase activator 1 (NOXA1) on serine 282 by MAP kinases and on serine 172 by protein kinase C and protein kinase A prevents NOX1 hyperactivation. *FASEB J.* 24, 2077–2092.
- (82) Chen, K., Kirber, M. T., Xiao, H., Yang, Y., and Keaney, J. F., Jr. (2008) Regulation of ROS signal transduction by NADPH oxidase 4 localization. *J. Cell Biol.* 181, 1129–1139.
- (83) Fridovich, I. (1995) Superoxide radical and superoxide dismutases. *Annu. Rev. Biochem.* 64, 97–112.
- (84) Gardner, P. R., Raineri, L., Epstein, L. B., and White, C. W. (1995) Superoxide radical and iron modulate aconitase activity in mammalian cells. *J. Biol. Chem.* 270, 13399–13405.
- (85) Juarez, J. C., Manuia, M., Burnett, M. E., Betancourt, O., Boivin, B., Shaw, D. E., Tonks, N. K., Mazar, A. P., and Donate, F. (2008) Superoxide dismutase 1 (SOD1) is essential for H₂O₂-mediated oxidation and inactivation of phosphatases in growth factor signaling. *Proc. Natl. Acad. Sci. U.S.A.* 105, 7147–7152.
- (86) DeYulia, G. J., Jr., Carcamo, J. M., Borquez-Ojeda, O., Shelton, C. C., and Golde, D. W. (2005) Hydrogen peroxide generated extracellularly by receptor-ligand interaction facilitates cell signaling. *Proc. Natl. Acad. Sci. U.S.A.* 102, 5044–5049.
- (87) DeYulia, G. J., Jr., and Carcamo, J. M. (2005) EGF receptor-ligand interaction generates extracellular hydrogen peroxide that inhibits EGFR-associated protein tyrosine phosphatases. *Biochem. Biophys. Res. Commun.* 334, 38–42.
- (88) Bienert, G. P., Schjoerring, J. K., and Jahn, T. P. (2006) Membrane transport of hydrogen peroxide. *Biochim. Biophys. Acta* 1758, 994–1003.
- (89) Bienert, G. P., Moller, A. L., Kristiansen, K. A., Schulz, A., Moller, I. M., Schjoerring, J. K., and Jahn, T. P. (2007) Specific aquaporins facilitate the diffusion of hydrogen peroxide across membranes. *J. Biol. Chem.* 282, 1183–1192.
- (90) Miller, E. W., Dickinson, B. C., and Chang, C. J. (2010) Aquaporin-3 mediates hydrogen peroxide uptake to regulate downstream intracellular signaling. *Proc. Natl. Acad. Sci. U.S.A.* 107, 15681–15686.

- (91) Wood, Z. A., Schroder, E., Robin Harris, J., and Poole, L. B. (2003) Structure, mechanism and regulation of peroxiredoxins. *Trends Biochem. Sci.* 28, 32–40.
- (92) Kang, S. W., Chae, H. Z., Seo, M. S., Kim, K., Baines, I. C., and Rhee, S. G. (1998) Mammalian peroxiredoxin isoforms can reduce hydrogen peroxide generated in response to growth factors and tumor necrosis factor- α . *J. Biol. Chem.* 273, 6297–6302.
- (93) Handy, D. E., Lubos, E., Yang, Y., Galbraith, J. D., Kelly, N., Zhang, Y. Y., Leopold, J. A., and Loscalzo, J. (2009) Glutathione peroxidase-1 regulates mitochondrial function to modulate redox-dependent cellular responses. *J. Biol. Chem.* 284, 11913–11921.
- (94) Halvey, P. J., Watson, W. H., Hansen, J. M., Go, Y. M., Samali, A., and Jones, D. P. (2005) Compartmental oxidation of thiol-disulphide redox couples during epidermal growth factor signalling. *Biochem. J.* 386, 215–219.
- (95) Winterbourn, C. C. (2008) Reconciling the chemistry and biology of reactive oxygen species. *Nat. Chem. Biol.* 4, 278–286.
- (96) Parsonage, D., Karplus, P. A., and Poole, L. B. (2008) Substrate specificity and redox potential of AhpC, a bacterial peroxiredoxin. *Proc. Natl. Acad. Sci. U.S.A.* 105, 8209–8214.
- (97) Peskin, A. V., Low, F. M., Paton, L. N., Maghzal, G. J., Hampton, M. B., and Winterbourn, C. C. (2007) The high reactivity of peroxiredoxin 2 with H_2O_2 is not reflected in its reaction with other oxidants and thiol reagents. *J. Biol. Chem.* 282, 11885–11892.
- (98) Woo, H. A., Yim, S. H., Shin, D. H., Kang, D., Yu, D. Y., and Rhee, S. G. (2010) Inactivation of peroxiredoxin I by phosphorylation allows localized H_2O_2 accumulation for cell signaling. *Cell* 140, 517–528.
- (99) Buhrow, S. A., Cohen, S., and Staros, J. V. (1982) Affinity labeling of the protein kinase associated with the epidermal growth factor receptor in membrane vesicles from A431 cells. *J. Biol. Chem.* 257, 4019–4022.
- (100) Clark, S., and Konstantopoulos, N. (1993) Sulphydryl agents modulate insulin- and epidermal growth factor (EGF)-receptor kinase via reaction with intracellular receptor domains: Differential effects on basal versus activated receptors. *Biochem. J.* 292 (Part 1), 217–223.
- (101) Woltjer, R. L., and Staros, J. V. (1997) Effects of sulphydryl modification reagents on the kinase activity of the epidermal growth factor receptor. *Biochemistry* 36, 9911–9916.
- (102) Seo, Y. H., and Carroll, K. S. (2009) Profiling protein thiol oxidation in tumor cells using sulfenic acid-specific antibodies. *Proc. Natl. Acad. Sci. U.S.A.* 106, 16163–16168.
- (103) Reddie, K. G., Seo, Y. H., Muse, W. B., III, Leonard, S. E., and Carroll, K. S. (2008) A chemical approach for detecting sulfenic acid-modified proteins in living cells. *Mol. Biosyst.* 4, 521–531.
- (104) Leonard, S. E., Reddie, K. G., and Carroll, K. S. (2009) Mining the thiol proteome for sulfenic acid modifications reveals new targets for oxidation in cells. *ACS Chem. Biol.* 4, 783–799.
- (105) Seo, Y. H., and Carroll, K. S. (2011) Quantification of Protein Sulfenic Acid Modifications Using Isotope-Coded Dimedone and Iododimedone. *Angew. Chem., Int. Ed.* 50, 1342–1345.
- (106) Truong, T. H., Garcia, F. J., Seo, Y. H., and Carroll, K. S. (2011) Isotope-coded chemical reporter and acid-cleavable affinity reagents for monitoring protein sulfenic acids. *Bioorg. Med. Chem. Lett.* 21, 5015–5020.
- (107) Leonard, S. E., and Carroll, K. S. (2011) Chemical 'omics' approaches for understanding protein cysteine oxidation in biology. *Curr. Opin. Chem. Biol.* 15, 88–102.
- (108) Benitez, L. V., and Allison, W. S. (1974) The inactivation of the acyl phosphatase activity catalyzed by the sulfenic acid form of glyceraldehyde 3-phosphate dehydrogenase by dimedone and olefins. *J. Biol. Chem.* 249, 6234–6243.
- (109) Saxon, E., and Bertozzi, C. R. (2000) Cell surface engineering by a modified Staudinger reaction. *Science* 287, 2007–2010.
- (110) Rostovtsev, V. V., Green, L. G., Fokin, V. V., and Sharpless, K. B. (2002) A stepwise Huisgen cycloaddition process: Copper(I)-catalyzed regioselective "ligation" of azides and terminal alkynes. *Angew. Chem., Int. Ed.* 41, 2596–2599.
- (111) Zhang, J., Yang, P. L., and Gray, N. S. (2009) Targeting cancer with small molecule kinase inhibitors. *Nat. Rev. Cancer* 9, 28–39.
- (112) Singh, J., Petter, R. C., and Kluge, A. F. (2010) Targeted covalent drugs of the kinase family. *Curr. Opin. Chem. Biol.* 14, 475–480.
- (113) Singh, J., Petter, R. C., Baillie, T. A., and Whitty, A. (2011) The resurgence of covalent drugs. *Nat. Rev. Drug Discovery* 10, 307–317.
- (114) Yun, C. H., Mengwasser, K. E., Toms, A. V., Woo, M. S., Greulich, H., Wong, K. K., Meyerson, M., and Eck, M. J. (2008) The T790M mutation in EGFR kinase causes drug resistance by increasing the affinity for ATP. *Proc. Natl. Acad. Sci. U.S.A.* 105, 2070–2075.
- (115) Yun, C. H., Boggon, T. J., Li, Y., Woo, M. S., Greulich, H., Meyerson, M., and Eck, M. J. (2007) Structures of lung cancer-derived EGFR mutants and inhibitor complexes: Mechanism of activation and insights into differential inhibitor sensitivity. *Cancer Cell* 11, 217–227.
- (116) Zhang, X., Gureasko, J., Shen, K., Cole, P. A., and Kuriyan, J. (2006) An allosteric mechanism for activation of the kinase domain of epidermal growth factor receptor. *Cell* 125, 1137–1149.
- (117) Jaffrey, S. R., Erdjument-Bromage, H., Ferris, C. D., Tempst, P., and Snyder, S. H. (2001) Protein S-nitrosylation: A physiological signal for neuronal nitric oxide. *Nat. Cell Biol.* 3, 193–197.
- (118) Jaffrey, S. R., and Snyder, S. H. (2001) The biotin switch method for the detection of S-nitrosylated proteins. *Sci. STKE* 2001, pl1.
- (119) Wang, H., and Xian, M. (2011) Chemical methods to detect S-nitrosation. *Curr. Opin. Chem. Biol.* 15, 32–37.
- (120) Giustarini, D., Dalle-Donne, I., Colombo, R., Milzani, A., and Rossi, R. (2008) Is ascorbate able to reduce disulfide bridges? A cautionary note. *Nitric Oxide* 19, 252–258.
- (121) Estrada, C., Gomez, C., Martin-Nieto, J., De Frutos, T., Jimenez, A., and Villalobo, A. (1997) Nitric oxide reversibly inhibits the epidermal growth factor receptor tyrosine kinase. *Biochem. J.* 326 (Part 2), 369–376.
- (122) Murillo-Carretero, M., Torroglosa, A., Castro, C., Villalobo, A., and Estrada, C. (2009) S-Nitrosylation of the epidermal growth factor receptor: A regulatory mechanism of receptor tyrosine kinase activity. *Free Radical Biol. Med.* 46, 471–479.
- (123) Lam, Y. W., Yuan, Y., Isaac, J., Babu, C. V., Meller, J., and Ho, S. M. (2010) Comprehensive identification and modified-site mapping of S-nitrosylated targets in prostate epithelial cells. *PLoS One* 5, e9075.
- (124) Glynn, S. A., Boersma, B. J., Dorsey, T. H., Yi, M., Yfantis, H. G., Ridnour, L. A., Martin, D. N., Switzer, C. H., Hudson, R. S., Wink, D. A., Lee, D. H., Stephens, R. M., and Ambs, S. (2010) Increased NOS2 predicts poor survival in estrogen receptor-negative breast cancer patients. *J. Clin. Invest.* 120, 3843–3854.
- (125) Switzer, C. H., Glynn, S. A., Cheng, R. Y., Ridnour, L. A., Green, J. E., Ambs, S., and Wink, D. A. (2012) S-Nitrosylation of EGFR and Src Activates an Oncogenic Signaling Network in Human Basal-Like Breast Cancer. *Mol. Cancer Res.* 10, 1203–1215.
- (126) Penel, S., Hughes, E., and Doig, A. J. (1999) Side-chain structures in the first turn of the α -helix. *J. Mol. Biol.* 287, 127–143.
- (127) Anderson, T. A., and Sauer, R. T. (2003) Role of an N_{cap} residue in determining the stability and operator-binding affinity of Arc repressor. *Biophys. Chem.* 100, 341–350.
- (128) Miranda, J. J. (2003) Position-dependent interactions between cysteine residues and the helix dipole. *Protein Sci.* 12, 73–81.
- (129) Kemble, D. J., and Sun, G. (2009) Direct and specific inactivation of protein tyrosine kinases in the Src and FGFR families by reversible cysteine oxidation. *Proc. Natl. Acad. Sci. U.S.A.* 106, 5070–5075.
- (130) Singh, J., Dobrusin, E. M., Fry, D. W., Haske, T., Whitty, A., and McNamara, D. J. (1997) Structure-based design of a potent, selective, and irreversible inhibitor of the catalytic domain of the erbB receptor subfamily of protein tyrosine kinases. *J. Med. Chem.* 40, 1130–1135.
- (131) Leonard, S. E., Garcia, F. J., Goodsell, D. S., and Carroll, K. S. (2011) Redox-based probes for protein tyrosine phosphatases. *Angew. Chem., Int. Ed.* 50, 4423–4427.